

# Microbial Contamination and Control Conference

## Endotoxin Deviations: A Practical Approach for Laboratory Investigators



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Eli Lilly and Company



# Microbial Contamination and Control Conference

May 8<sup>th</sup> & 9<sup>th</sup>



The information to follow are examples from real deviation investigations but all recommendations, mitigation strategies, and points to consider are my own opinion and not that of Eli Lilly & Co.



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## Agenda

1. Background
2. Initial Lab Investigation
3. Investigation Testing
4. OOS Handling & Approach
5. Overcoming Inhibition / Enhancement
6. Case Studies
7. System Suitability Issues & Mitigations
8. Conclusion





## PDA Tech Report 88 – Micro Deviations

- Phase I – Lab Investigation or Analytical Investigation
  - QC / Lab Management / QA / SMEs
  - Goal – Establish validity of atypical result & determine if lab assignable cause or not
- Phase II – Manufacturing Investigation
  - Sterility Assurance / Ops / Engineering / QA / QC SME / Management
  - Goal – Determine if at any part of the manufacturing process could have led to atypical result

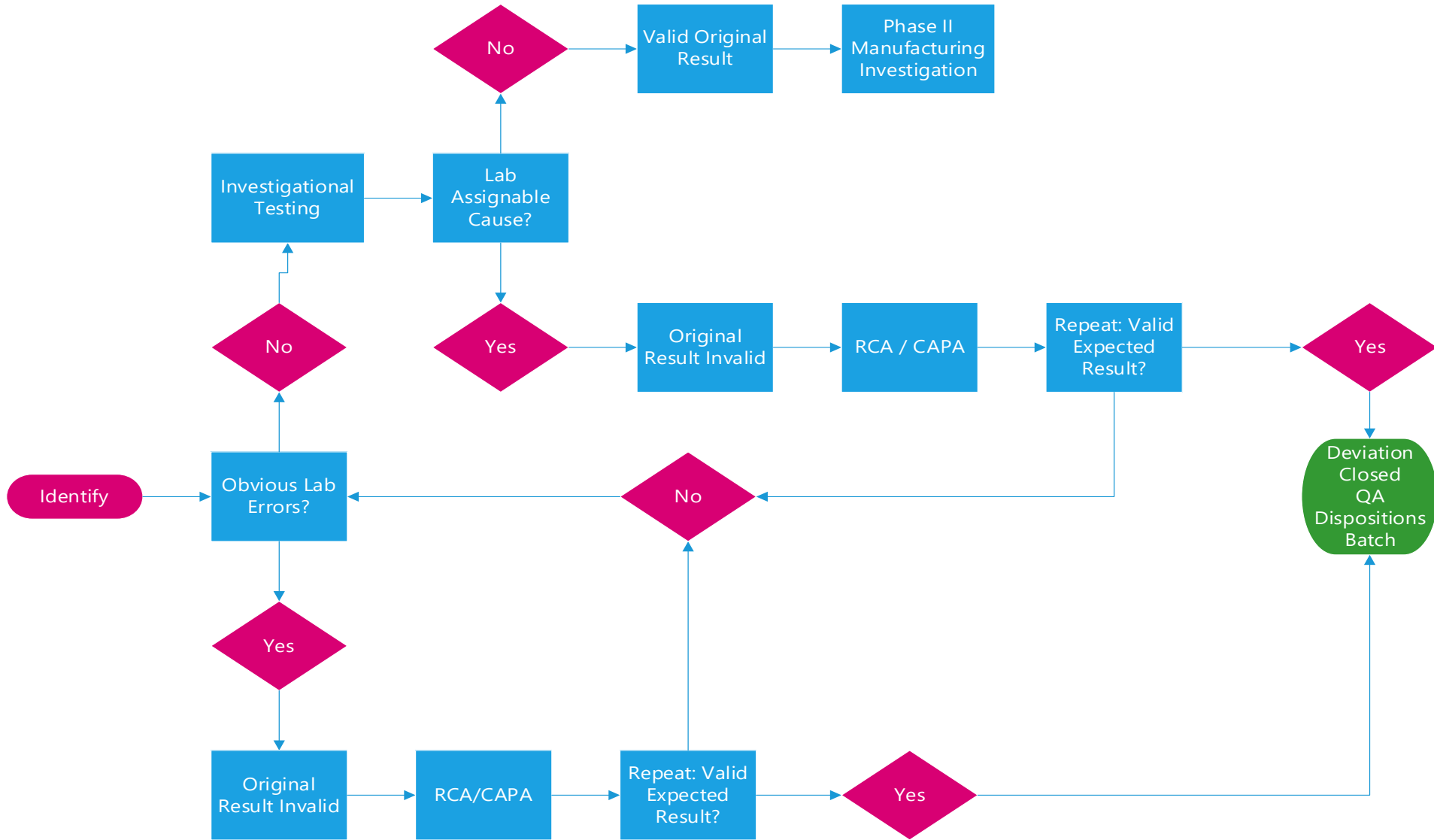


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## Flow Diagram of Phase I Endotoxin Investigation



\*Goal → Determine Validity & Root Cause



## Obvious Lab Errors



### Sample Preparations

- Save dilutions until sample release
- Not knowingly continue a test expected to be invalidated later
- Right the First Time Culture



### Reagents & Consumables

- Shown to be free of endotoxin & noninterfering
- Label claim qualification each shipment standards



### Instrument

- Audit trail reviews
- Roles & responsibilities defined & controlled



### Analyst

- Training & Qualification
- Analyst interview



# Interview

Which question will lead to a better dialogue and nuances of method execution:

“Did you follow sample preparation described in the analytical method?”

“Describe how you executed sample preparation for the analytical method.”

Practice vs Procedure gaps, lack of knowledge or training



## Conducting Interviews

### • DO

- Write questions down
- Ask open ended questions
- Ask for input from the interviewee
- Stimulate back and forth conversation
- Start with open ended questions then narrow down to specific yes or no



### • DON'T

- Ask leading questions
- Assume or place blame
- Forget to ask for feedback on the process, method, procedure, etc







## Investigational Testing for BET

### Determine Validity

- Assume results are valid until proven otherwise

### Formulate and test hypothesis

- Confirm or discount
- Not repeat – cannot be used as final result

### BET

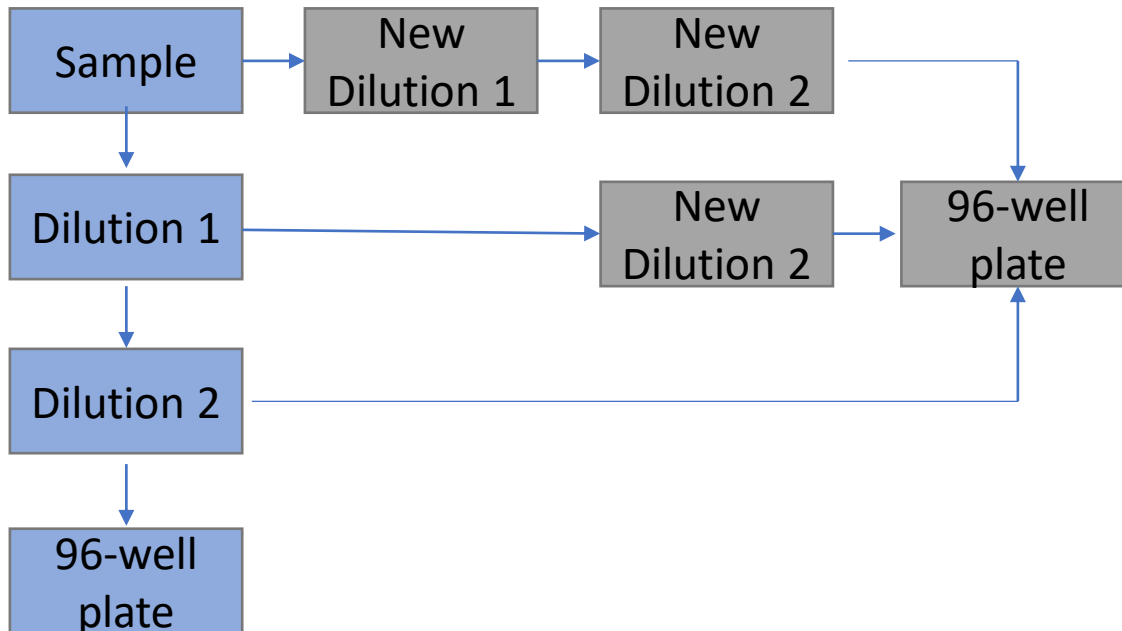
- Testing dilution tubes – contamination introduced during prep
- Repeat of original – 96-well plate suspected or instrument error
- New standard preparation
- New reagent preparation
- pH



# Investigational Testing

- Reagents (LRW) → passing Negative Control
- Hypothesis → Contamination during sample prep
- Investigational Testing → Confirm or Disprove Hypothesis
- Outcome → Hypothesis confirmed

| Samples        | Dilution | Well             | Reaction Time (sec) | Average Reaction Time (sec) | Raw EU                                       | Results (Linear Regression) EU/mL | Release Limit |
|----------------|----------|------------------|---------------------|-----------------------------|--|-----------------------------------|---------------|
| S1             | 1        | A2               | 2784                | 2813                        | 0.0357                                       | 0.0357                            | N/A           |
|                |          | B2               | 2841                |                             |  |                                   |               |
| PPC            | 1        | C2               | 1884                | 1926                        | 0.141  |                                   |               |
|                |          | D2               | 1967                |                             |  |                                   |               |
| PPC Value: 0.1 |          | % PPC Recovery : |                     | 105%                        | (PPC - SAMPLE 1) Endotoxin Recovered : 0.105 |                                   |               |



| Outcome  |               |
|--|---------------|
| <b>New Dilution 2 from Original Sample</b>     | <0.0100 EU/mL |
| <b>New Dilution 2 from Original Dilution 1</b> | 0.0282 EU/mL  |
| <b>Original Dilution 2</b>                     | 0.0230 EU/mL  |



## Out of Specification Handling & Approach

Establish Validity

Even invalid OOS should be rigorously investigated

Cannot invalidate without lab assignable cause

5x Retest

Regulatory Experience

Provide list of valid and invalid OOS investigations



## OOS Investigational Testing

7 excipient samples with endotoxin activity resulting in an OOS event

| Obvious lab errors                             | Reagents  | Investigational Testing   | Isolated Event?   |
|--|---|---|---|
| Sample preparation<br>Consumables<br>Interview | Negative Controls<br>Common reagent<br>50/50 v/v<br>Dispersing/Buffer | 50mM Buffer<br>→ <0.01EU/mL<br>Dispersing Reagent Vial<br>→ 0.13EU/mL<br>0.5% Dispersing Reagent<br>→ 0.12EU/mL<br>50/50 Dispersing/Buffer<br>→ 0.05EU/mL | Vials before and after event<br>Vendor inquiry<br>Analyst coaching<br>Switch lots |



## Overcoming Inhibition / Enhancement

pH  
adjustment

Buffers

Dispersing  
agents

Mixing

MVD





## MVD

- $MVD = (\text{endotoxin limit} \times \text{sample concentration}) / \lambda$
- Guidance in <85>, <1085>, proposed <86>
  - Dilute to MVD
- Pooled samples adjustment
  - For example: MVD is 1500 but 3 vials are pooled  $\rightarrow$  1/3 MVD is 500



## Material of Construction

Regulatory focus during inspections

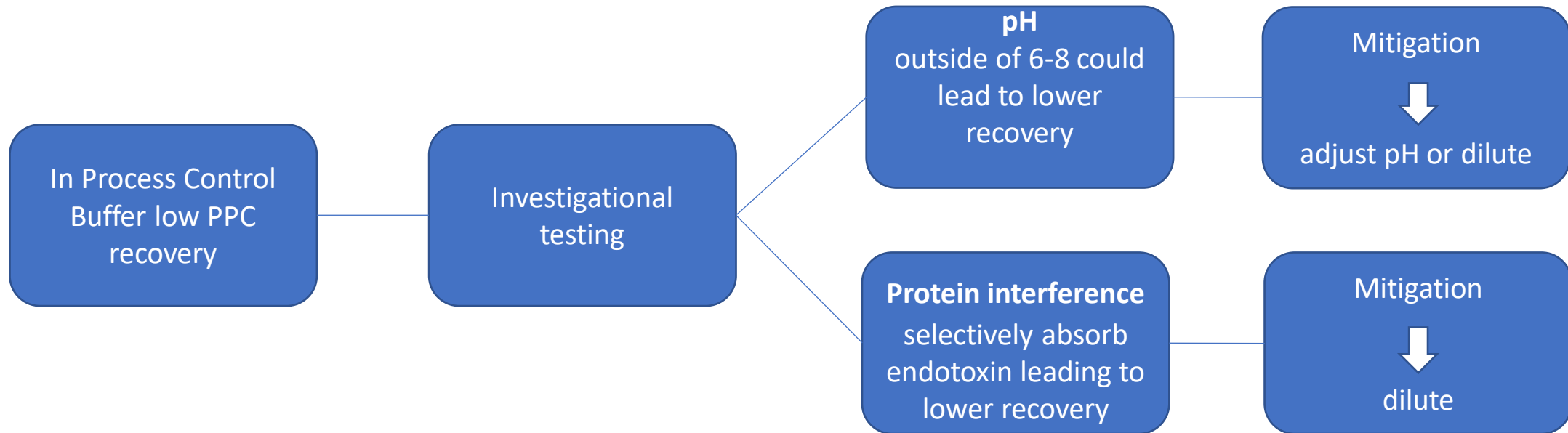
Guidance from 2012 FDA Testing Questions & Answers Document  
Established Hold Times  
PETG, PS, PE  
EVA, ULDPE  
Do not use PP  
QC Labs need to be looped in manufacturing changes impacting sample containers

LER

Included in BLA submissions  
Focus on matrix interaction  
i.e. chelator and surfactant in the matrix



## Low PPC Deviation Investigation



- pH: 7.52 → within recommended range
- UV analysis → positive for protein content
  - Original sample was diluted 1:10
  - IPC DP requires 1:100

- Manufacturing Investigation (Phase II)
  - Confirmed sample pulled from wrong tank



## Low PPC Deviation Investigation

### Original Verification

- Single supplier of rFC Reagent
- Diluent – MgCl<sub>2</sub>

### Low PPC Recovery

- Secondary supplier utilized
- On going trend with material

### Follow up method development activities

- Additional testing confirmed MgCl<sub>2</sub> interference with 2<sup>nd</sup> supplier
- Updated diluent and dilution scheme to work with both suppliers
- Tris Buffer

| Supplier                            | Avg. PPC Rec (%) | Number of Samples |
|-------------------------------------|------------------|-------------------|
| Supplier 1 Verification             | 97               | 3                 |
| Supplier 1                          | 96               | 17                |
| Supplier 2                          | 51               | 4                 |
| Supplier 2 Post Verification Update | 91               | 19                |

**2<sup>nd</sup> supplier verification on all commercialized molecules utilizing rFC platform**

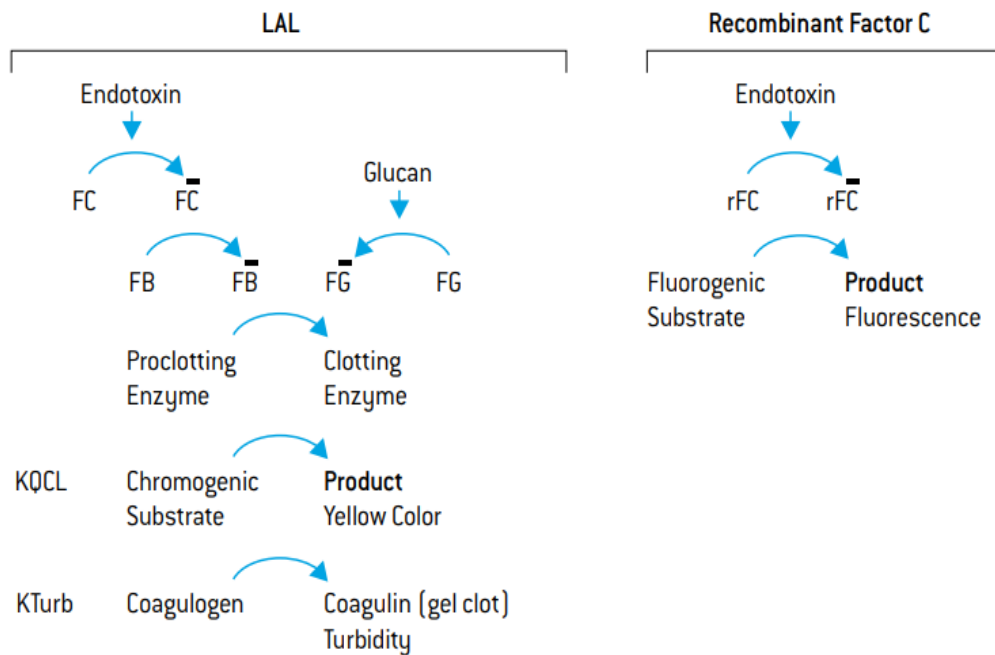


## Beta Glucan Interference

### Beta glucan interference

- Activate factor G pathway, false positive
- cellulose filter in the manufacturing process
- raw bulk materials (yeastolate)

Recombinant assays mitigate beta glucan false positive interference in LAL assays



\*Courtesy of Lonza package insert.

| Supplier   | Reagent     | Sample     | Result (EU/mg) | %PPC Recovery |
|------------|-------------|------------|----------------|---------------|
| Supplier 2 | LAL         | Yeastolate | 0.0687         | 133           |
| Supplier 3 | LAL         |            | >4.00          | N/A           |
| Supplier 6 | LAL         |            | 0.0639         | 123           |
| Supplier 1 | Recombinant |            | <0.0400        | 79            |
| Supplier 2 | Recombinant |            | <0.0400        | 71            |
| Supplier 3 | Recombinant |            | <0.0400        | 100           |
|            | Recombinant |            | <0.0400        | 87            |
| Supplier 4 | Recombinant |            | <0.0400        | 56            |
| Supplier 5 | Recombinant |            | <0.0400        | 76            |
|            | Recombinant |            | <0.0400        | 91            |
| Supplier 6 | Recombinant |            | <0.0200        | 107           |





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## USP <86> if approved, early adoption in Nov 2024

### Recombinant chapter

- rFC – end point florescence
- rCR – chromogenic, absorbance



### Compendia Impact

- PhEur - Replacing RPT with MAT – 2026
- Could LAL be next?



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## Case Study #1 – Cleaning Validation Samples



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## Background

- Submitted as WFI
- Analyzed on alternate rFC platform
- Controls passed
- Rinse samples inhibited

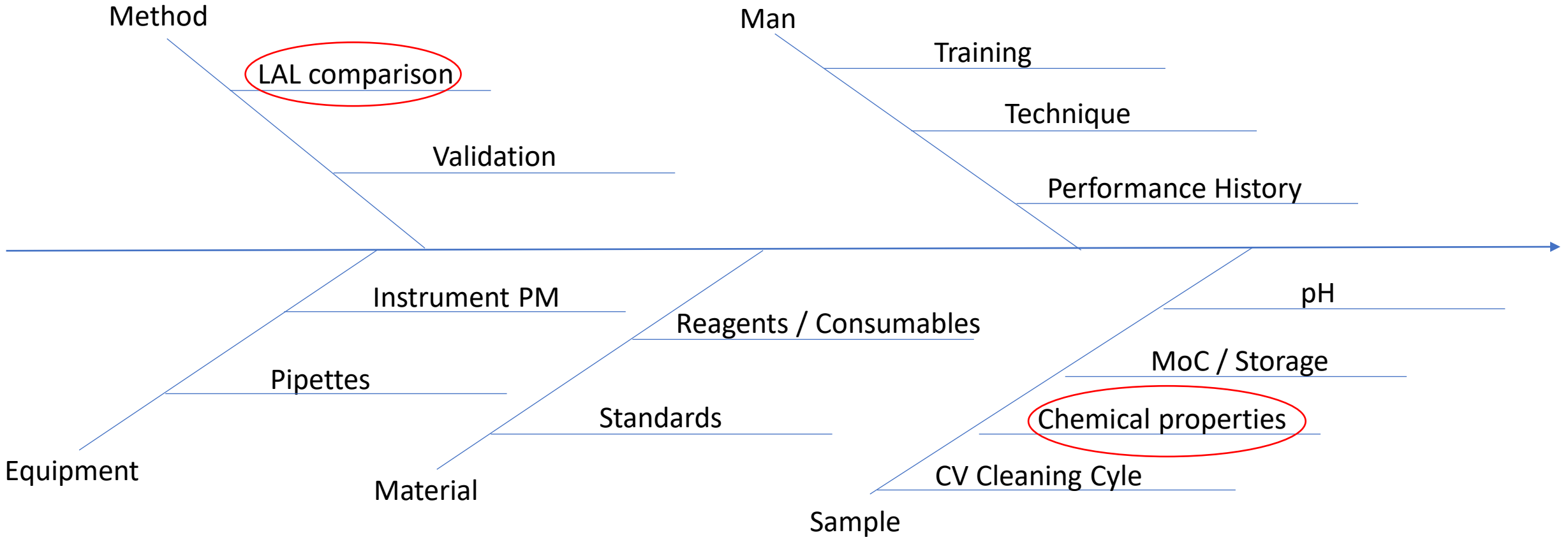
## Impact

- Multiple deviations
- Manufacturing equipment



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# Initial Analytical Investigation



pH of sample + rFC reagent



pH of control & rinse sample



Normal results:

- Particulates
- Bioburden
- Phosphate
- Conductivity



Business Continuity Plan - LAL





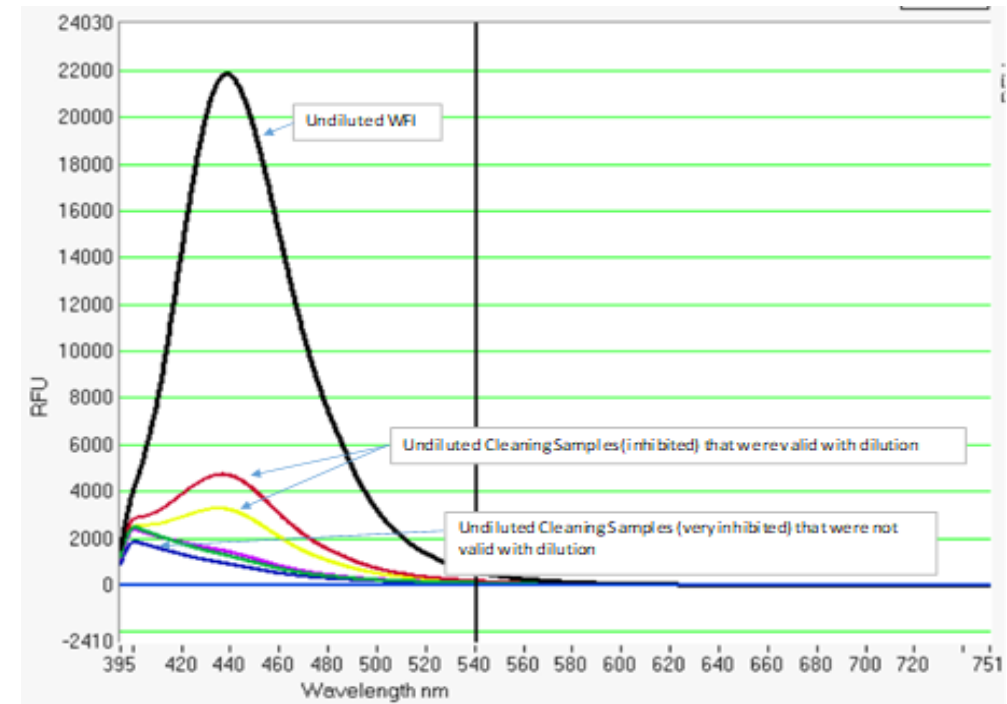
## Detailed Investigation

### Assumption was rinse sample is equivalent to WFI

- Residual product or cleaning reagent?
- Cleaning cycle passed indicating the appropriate removal of cleaning agent and any residual product

### Investigation testing

- Spectral scans: submitted cleaning samples  $\neq$  WFI
- Trace amounts of CIP (surfactant) inhibit PPC recovery on rFC and not LAL
- Certain reagent suppliers more sensitive to interference than others





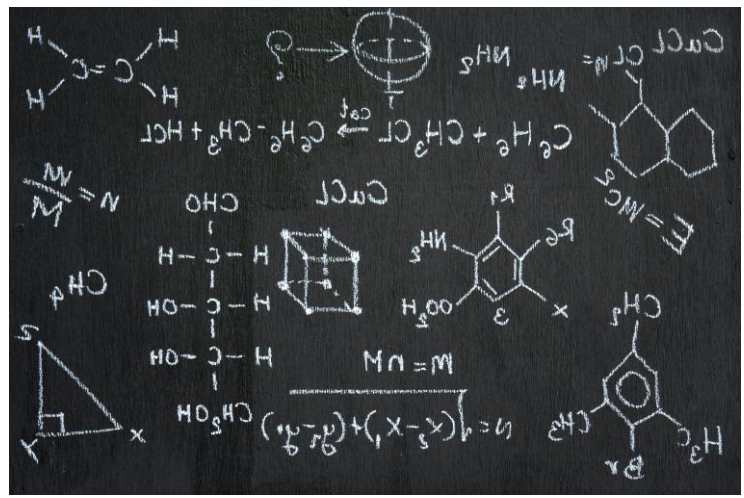
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## CAPA

- Method Development
- Harmonized dilution scheme



## Learning

- Consider all impacts when evaluating process change or implementing new methodologies / technologies
- Method was optimized for WFI, Clean Steam
- Assumption was rinse water = WFI



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## Case Study #2 – Plate Reader PM Failure



## Background & Impact

Vendor PM

Uniformity on Fluorescence Readers

Absorbance Readers

“Tagged Out” Plate Readers

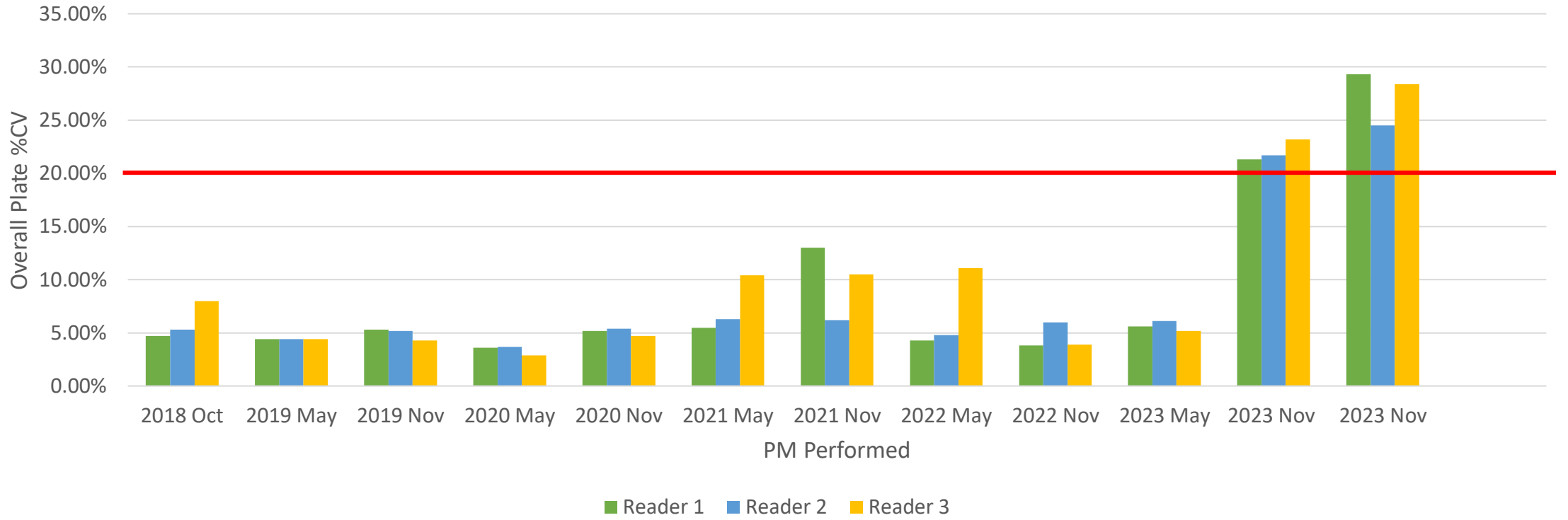


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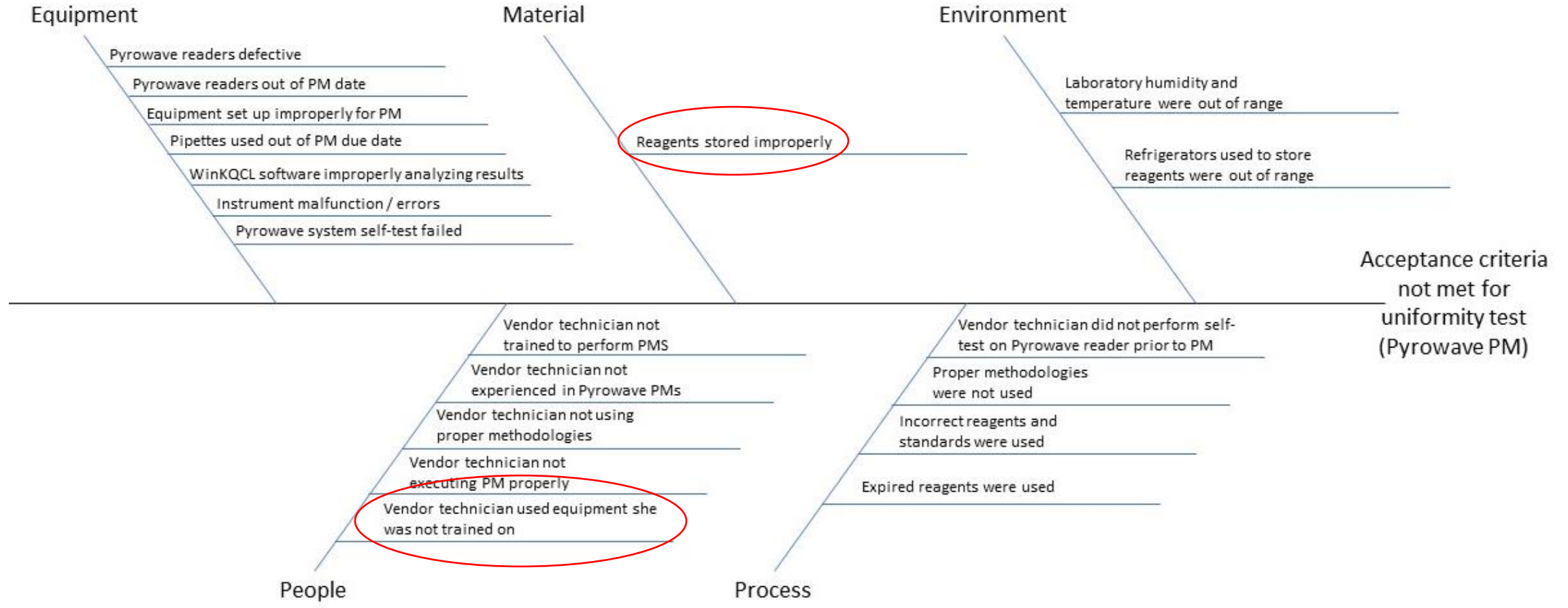
Historical Results  
October 2018 – November 2023





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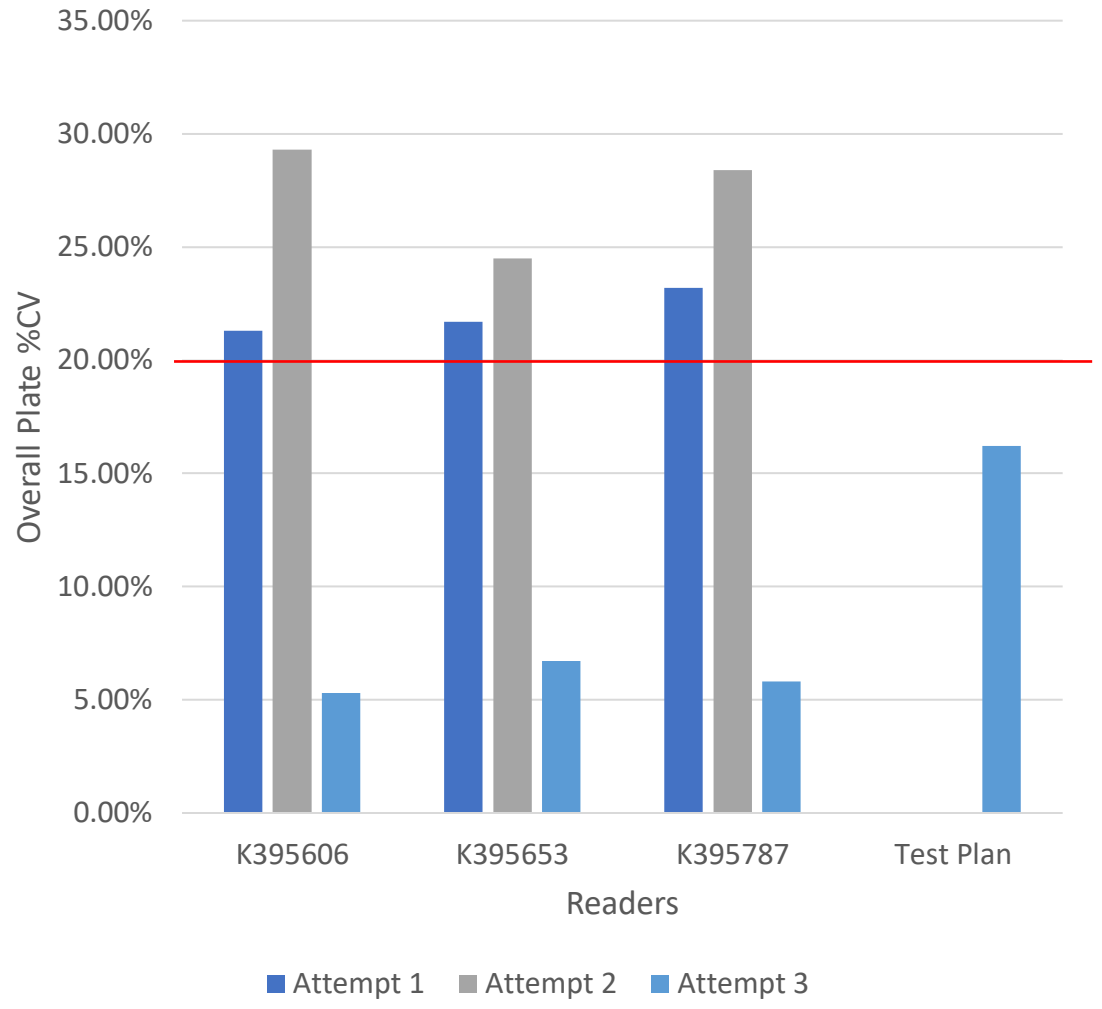






## Investigational Testing

- Properly stored reagents
  - %CV = 16.2%
- Familiar Pipettes
  - Attempt 3





## CAPA & Summary

### Impact:

- Plate Reader PM Uniformity Test failures on all 3 Micro QC lab Readers
- 2 weeks

### Root Causes:

- Improper storage of reagents
- Vendor technician using equipment without training

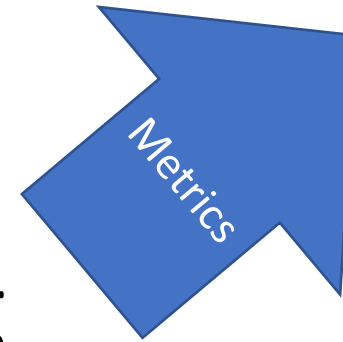
### CAPA:

- Procedural updates
- Network Shared Learning

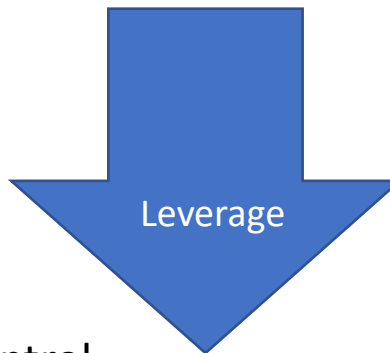


## System Suitability Issues

“Invalid test should be tracked and trended to look for patterns and trends that might require a corrective action”



Trending



- Invalid assay rate
- Invalid sample rate
- %PPC Recovery
- %CV for PPC & Standard wells

How does this look for our lab?

- Document every system & sample control issue
- Monthly tracking of metrics
- Upper control limit established

- State of Control
- Support repeat testing without Deviation record



# Mitigation Considerations



## Training

- Robust Training Program
- Observation & Hands on
- Intervention for identified trends



## Ready to Use Plates

- Minimize analyst technique issues & pipette variations at small volumes



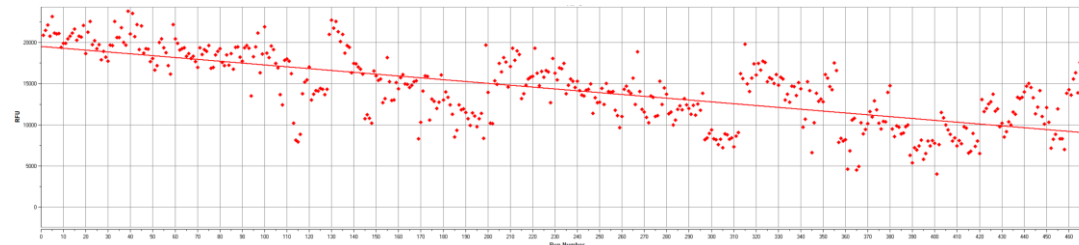
## Reagent & Standards

- Monitoring standard signal / response (end point fluorescence) & reaction times (Kinetic LAL)



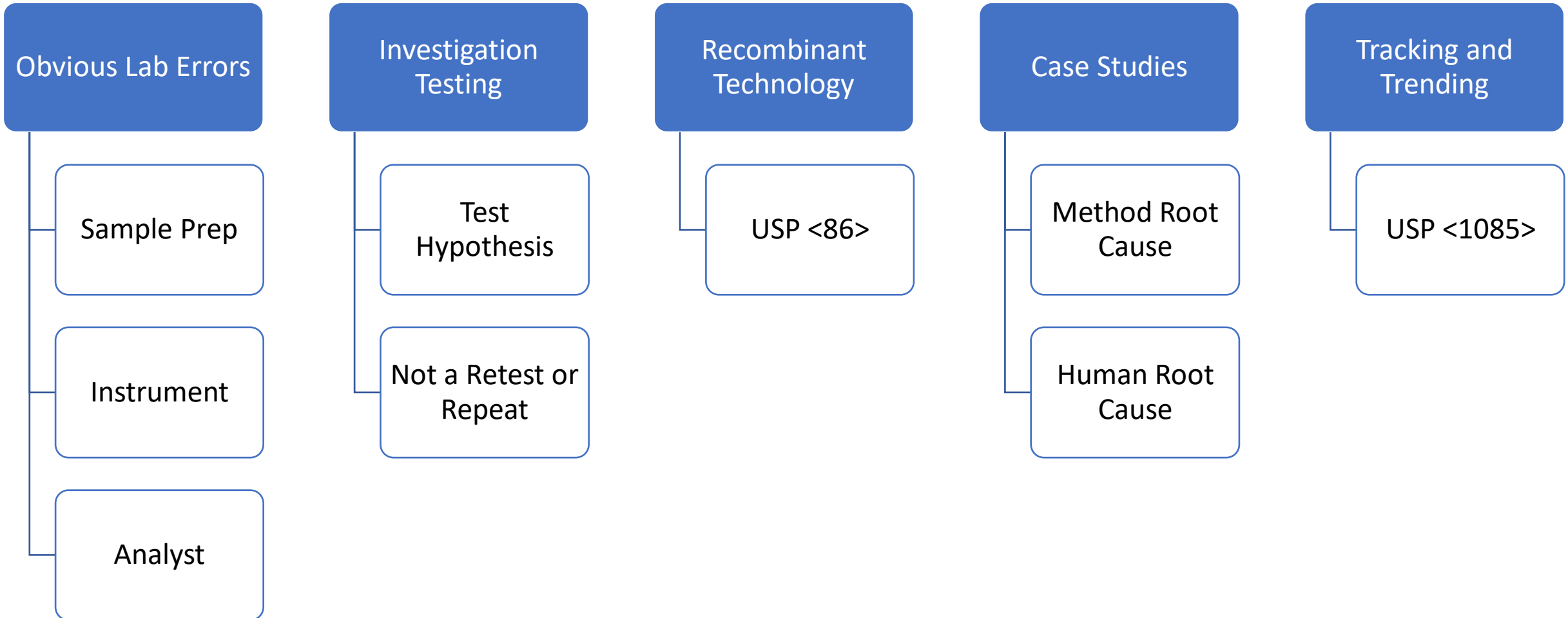
## Instrument Optimization (Fluorescent Readers)

- Scan rate
- Sensitivity / Gain
  - Utilize entire dynamic range





## Conclusion





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# Questions?